



FEMOBIOME® AlphaScreen FEMOBIOME® DeltaScreen

Line of new tests for women microbiota assessment

## Content

What's new?	3
Comparison of FEMOBIOME® tests	
How to choose test?	6
Types of biomaterial	7
Preparation and sample collection	7
New test report	11
Main technological features	12

## + What's new?

Expanded and optimized panel of detectable microorganisms enables to provide clinically significant description of microbiota composition

More informative evaluation of normobiota state for women of different ages;

Quantitative assessment of Group B Streptococcus – especially important during pregnancy planning and gestation;

Control of obligate pathogens presence to exclude asymptomatic and subclinical infections:

Detection of viruses relevant to reproductive health.

System of visual markers for convenient clinical interpretation

Test report with personalized comments and infographics;

New types of biomaterial

Cervical swabs collected in transport medium for liquid-based cytology;

Self-collected vaginal samples

Expanded line of FEMOBIOME® test kits

FEMOBIOME II – universal test for assessment women microbiota state with the most comprehensive capabilities;

FEMOBIOME\*AlphaScreen – a shortened version of FEMOBIOME\*II for diagnostics of bacterial vaginosis;

FEMOBIOME\*DeltaScreen – a complex test for diagnostics of infectious and inflammatory urogenital diseases.



## 

	FEMOBIOME°				
	FB°16	FB°II	Screen	Delta Screen	Alpha Screen
Number of tubes	16	16	8	8	8
Indicators					
Human genomic DNA	•	•	•	•	•
Total bacterial load	•	•	•	•	•
Normobiota					
Lactobacillus spp.	•	•	•	•	•
L. crispatus		•			
L. jensenii/ L. mulieris		•			
L. gasseri/ L. paragasseri		•			
L. iners		•		•	•
L. non-iners				•	•
Bifidobacterium spp.		•			
Aerobes					
Staphylococcus spp.	•	•		•	
Streptococcus spp.	•	•		•	
Streptococcus agalactiae		•			
Enterobacteriaceae	•	•		•	
Enterococcus spp.		•		•	
Haemophilus spp.		•		•	
Anaerobes					
Gardnerella vaginalis	•	•	•	•	•
Fannyhessea vaginae (Atopobium vaginae)	•	•		•	•
Mobiluncus spp.	•	•		•	•
Anaerococcus spp.		•			•
Peptostreptococcus spp.	•	•	<u> </u>	<u> </u>	•

Bacteroides spp./Porphyromonas spp./Prevotella spp.	•	•			•
Sneathia spp./Leptotrichia spp. /Fusobacterium spp.	•	•			•
Megasphaera spp./ Veillonella spp./Dialister spp.	•	•			•
BVAB1 / BVAB2 / BVAB3		•			•
Mycoplasmas					
Ureaplasma urealyticum	spp.	•	spp.	•	•
Ureaplasma parvum	•	•	•	•	•
Mycoplasma hominis	•	•	•	•	•
Yeast fungi					
Candida spp.	•	•	•	•	•
Candida albicans		•		•	•
SEXUALLY TRANSMITTED INFECTIONS (	STI)				
Chlamydia trachomatis		•	•	•	
Mycoplasma genitalium	•	•	•	•	
Neisseria gonorrhoeae		•	•	•	
Trichomonas vaginalis		•	•	•	
Herpesviruses					
HSV-1		•	•	•	
HSV-2		•	•	•	
CMV		•	•	•	
Human papillomaviruses					
HPV 16		•			
HPV 18		•			
HPV 45		•			
HPV 31/ 33/ 35/ 39/ 51/ 52/ 56/ 58/ 59/ 66/ 68		•			

# FEMOBIOME\*II – more clinically relevant information for the same price

## ?

## How to choose test?

Clinical situation	FEMOBIOME°				
	FB°16	FB°II	Screen	Delta Screen	Alpha Screen
Infectious disease	es of repro	ductive s	ystem		
Inflammatory symptoms	•	•	•	•	
Symptoms of BV	•	•			•
Asymp	otomatic c	ases			
Preparation for pelvic surgery	•	•	•	•	
Preventive screening	•	•	•	•	
Treatm	ent monit	oring			
Treatment of infection diseases	•	•	•	•	
BV treatment	•	•			•
Obstetrics					
Pregnancy planning, ART	•	•	•	•	
Prophylactic examinations during pregnancy	•	•	•	•	
Infertility, miscarriage	•	•	•	•	

## Types of biomaterial

Vaginal swab. The sample can be collected by a healthcare specialist or self-collected by woman using special devices.

Cervical swab collected in transport medium, including transport medium for liquid cytology.



## Sample collection and preparation

### Common recommendations

PCR analysis is a direct method of laboratory research; therefore, the biological material needs to be collected from the foci of infection. The biomaterial type is selected by the physician based on the collected anamnesis and clinical picture of the disease.

The quality of sampling, sample storage, transport and pretreatment are crucial for obtaining correct results. Incorrect sampling may lead to unreliable results and, therefore, to the need to repeat the sampling procedure.

Sample collection is performed using specialized medical devices.

After sample collection put the swab into the tube with transport medium and rinse it thoroughly. Remove the swab from solution, press it to the wall of tube and squeeze the rest of the liquid.

Throw out the swab. Use a new swab if you need to repeat sampling or to take sample from another biotope.

### Sample collection and preparation

Women must not perform hygiene procedures or syringing prior the sampling procedure. To interpret results successfully and robustly, sample must contain the largest possible number of epithelial cells with minimum amounts of mucus and blood. Before obtaining an epithelial cell swab from the posterior vaginal vault and cervical canal, the free-flowing secretion should be removed with a sterile cotton swab.

! Most microorganisms are localized on the epithelial surface in the form of biofilm. Therefore, the sample should include a scaring of the epithelial surface with adherent microbial layer, not the contents of the organ lumen.

BEFORE THE TEST, IT IS NOT ALLOWED TO	IF THE GUIDELINES ARE NOT FOLLOWED, biomaterial can be collected no sooner than
Perform syringing and vaginal douching with antiseptics	
Use tampons	
Use PCR-inhibiting substances, such as ultrasound gel, heparin, chlorhexidine and other chlorine- containing medicinal products	24 hours
Undergo transvaginal ultrasound examination	
Engage in protected sexual contacts	

Undergo colposcopy	48 hours		
Engage in unprotected sexual contacts	72 hours		
Take and/or use antibacterial	2 weeks		
medications, probiotics	after the therapy		
	4 weeks		
	after the therapy of Neisseria gonorrhoeae, Trichomonas vaginalis, Chlamydia trachomatis, Mycoplasma genitalium		

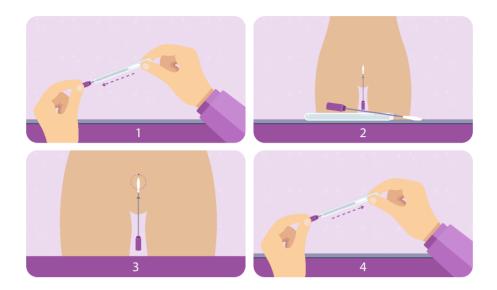
The test can be prescribed to patients on antibiotic therapy (for instance, for therapy monitoring purposes). However, antibiotics can influence the outcome of the test. Test results should be interpreted by the doctor based on the purpose of testing and the patient's medical history.

## Vaginal sampling

The sample must be taken prior to physical examination. Speculum can be treated with warm water before the procedure. Antiseptics and oil-based lubricants (like Vaseline) must not be used for speculum treatment. The sample must be taken from the lateral or posterolateral vaginal wall.

### Self-sampling

Self-sampling is performed using specialized medical devices Follow the manufacturer's instructions for correct use.



- 1 Open the package and remove the swab
- Take a comfortable position and insert the swab into the vagina up to the marked line
- 3 Rinse the swab for 10-30 second
- Remove the swab and place it into the sterile container provided in the kit
- 5 Deliver the sample to the laboratory

## Cervical sampling

Insert the swab 0,5–1,5 cm into the cervical canal.

Avoid any contact with the vaginal walls.



### Indicator icon color code – traffic light system

	PATHOGENS			
	Yes	No		
		Mycoplasmas* ≥ 4 lg GE/ml		
		Candida** ≥ 4 lg GE/ml		
		Yes	No	
Eubiosis	<b>(a)</b>		$\bigcirc$	
Moderate dysbiosis	<b>(a)</b>			
Severe dysbiosis				

<sup>\*</sup> Ureaplasma urealyticum Ureaplasma parvum Mycoplasma hominis

- Indicator icon visual summary of the microbiota status and presence of pathogens, based on the traffic light system (see table below)
- Linear histogram a graphical representation of the proportion of normobiota, anaerobes, aerobes and mycoplasmas in the microbiota composition. Color indicate the total quantity of microorganisms in each group
- Columnar histogram microorganisms profile showing detailes distribution of conditionally pathogenic microorganisms within their groups

<sup>\*\*</sup> Candida spp. Candida albicans

## Main technological features

All-in-one solution – from sample collection to interpretation the test report;

Two PCR setup formats – automated (384-well) and manual (96-well);

Fast nucleic acid extraction, optimized for Allsheng Auto-Pure 96 and KingFisher Flex (PREP-MB-RAPID II);

#### Biomaterial

epithelial swab from the mucous membrane of cervical canal, vagina

### Transport medium

STOR-F

STOR-M

### Transport medium for liquid cytology

PreservCyt<sup>®</sup>, Hologic Inc., USA

BD SurePath™ Liquid-Based Pap Test, Becton, Dickinson and Company, USA;

EASYPREP, YD Diagnostics, South Korea;

CellPrep, CP Biodyne, South Korea;

Cell Preservative Solution, Human Lituo Biotechnology Co.Ltd, China

### Sample pretreatment

PREP-PK (for scrapes taken in the transport & fixation medium for liquid cytology BD SurePath™ Liquid-Based Pap Test and CellPrep)

### Time of analysis

From 2,5 hours (with sample pretreatment)

## Number of samples

12 (24)\*\* tests for package S, including control samples

24 (48)\*\* tests for package A and A-TL, including control samples

## Storage conditions

+2 °C ... 8 °C – packages S, A, A-TL

-18 °C ...-22 °C – TechnoTaq MAX polymerase (part of package A)

12 months

<sup>\*\*</sup>for FEMOBIOME®AlphaScreen and FEMOBIOME®DeltaScreen

